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## Phosphorylation of BCL-2 regulates ER $\text{Ca}^{2+}$ homeostasis and apoptosis

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Phosphorylation of BCL-2 within an unstructured loop inhibits its antiapoptotic effect. We found that phosphorylated BCL-2 predominantly localized to the endoplasmic reticulum (ER) and tested whether phosphorylation would control its activity at this organelle, where  $\text{Ca}^{2+}$  dynamics serve as a critical control point for apoptosis. Phosphorylation greatly inhibits the ability of BCL-2 to lower  $[\text{Ca}^{2+}]_{\text{ER}}$  and protect against  $\text{Ca}^{2+}$ -dependent death stimuli. Cells expressing nonphosphorylatable BCL-2<sup>AAA</sup> exhibited increased leak of  $\text{Ca}^{2+}$  from the ER and further diminished steady-state  $[\text{Ca}^{2+}]_{\text{ER}}$  stores when compared to cells expressing BCL-2<sup>wt</sup>. Consequently, when BCL-2 is phosphorylated,  $\text{Ca}^{2+}$  discharge from the ER is increased, with a secondary increase in mitochondrial  $\text{Ca}^{2+}$  uptake. We also demonstrate that phosphorylation of BCL-2 inhibits its binding to proapoptotic family members. This inhibitory mechanism manifested at the ER, where phosphorylated BCL-2 was unable to bind proapoptotic members.  $[\text{Ca}^{2+}]_{\text{ER}}$  proved coordinate with the capacity of BCL-2 to bind proapoptotic BH3-only members, further integrating the apoptotic pathway and  $\text{Ca}^{2+}$  modulation. Unexpectedly, the regulation of ER  $\text{Ca}^{2+}$  dynamics is a principal avenue whereby BCL-2 phosphorylation alters susceptibility to apoptosis.

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possess sequence conservation within four domains (BH1-4), and bind and sequester proapoptotic proteins. Proapoptotic BCL-2 family proteins can be subdivided into 'multidomain' proteins such as BAX and BAK, which display homology within BH1-3 domains, and the 'BH3-only' proteins, which only possess sequence homology within this amphipathic  $\alpha$ -helix, which serves as the critical death domain. The ratio between the antiapoptotic and proapoptotic BCL-2 family members helps determine the susceptibility of cells to a death signal (Oltvai *et al.*, 1993). Genetic and biochemical studies indicate that multidomain BAX and BAK function as a gateway to the intrinsic death pathway operating at both the mitochondria and endoplasmic reticulum (ER). The upstream BH3-only members respond to select death signals and subsequently trigger the conformational activation of BAX and BAK, inducing their intramembranous homo-oligomerization and permeabilization of the mitochondrial outer membrane (Wei *et al.*, 2001). Released intermembranous proteins include cytochrome *c*, which complexes with Apaf-1 and caspase-9 to form a postmitochondrial apoptosome that activates effector caspases (Li *et al.*, 1997). Conversely, BCL-2 can sequester activated BH3-only proteins, and thus inhibit the activation of BAX and BAK (Cheng *et al.*, 2001).

In the absence of BAX and BAK, cells are resistant to a wide variety of death signals including agents that release  $\text{Ca}^{2+}$  from intracellular stores, such as the lipid second messengers arachidonic acid and  $\text{C}_2$ -ceramide as well as oxidative stress. BAX and BAK also localize to the ER, and cells deficient in BAX, BAK have reduced resting  $[\text{Ca}^{2+}]_{\text{ER}}$ , which accounts for their resistance to  $\text{Ca}^{2+}$ -dependent death stimuli (Scorrano *et al.*, 2003). Reciprocally, expression of the antiapoptotic protein BCL-2 protects cells from death by thapsigargin, an irreversible inhibitor of the sarcoplasmic-endoplasmic reticulum  $\text{Ca}^{2+}$  ATPase (SERCA) responsible for uptake of  $\text{Ca}^{2+}$  from the cytosol into the ER lumen (Lam *et al.*, 1994). BCL-2 is also found at the ER (in addition to mitochondria and nuclear membrane), and its overexpression resulted in reduced  $[\text{Ca}^{2+}]_{\text{ER}}$  accompanied by an increased

5. Induzione di proteine ricombinanti in colture di E. coli trasformate
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8. Il candidato legga e traduca in Italiano il primo paragrafo del seguente abstract

## Parkinson's disease mutations in PINK1 result in decreased Complex I activity and deficient synaptic function

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Mutations of the mitochondrial PTEN (phosphatase and tensin homologue)-induced kinases (PINK1) are important causes of recessive Parkinson disease (PD). Studies on loss of function and overexpression implicate PINK1 in apoptosis, abnormal mitochondrial morphology, impaired dopamine release and motor deficits. However, the fundamental mechanism underlying these various phenotypes remains to be clarified. Using fruit fly and mouse models we show that PINK1 deficiency or clinical mutations impact on the function of Complex I of the mitochondrial respiratory chain, resulting in mitochondrial depolarization and increased sensitivity to apoptotic stress in mammalian cells and tissues. In *Drosophila* neurons, PINK1 deficiency affects synaptic function, as the reserve pool of synaptic vesicles is not mobilized during rapid stimulation. The fundamental importance of PINK1 for energy maintenance under increased demand is further corroborated as this deficit can be rescued by adding ATP to the synapse. The clinical relevance of our observations is demonstrated by the fact that human wild type PINK1, but not PINK1 containing clinical mutations, can rescue Complex I deficiency. Our work suggests that Complex I deficiency underlies, at least partially, the pathogenesis of this hereditary form of PD. As Complex I dysfunction is also implicated in sporadic PD, a convergence of genetic and environmental causes of PD on a similar mitochondrial molecular mechanism appears to emerge.