



Course unit English denomination	Emerging Infectious diseases: pathogenesis, treatment and prevention
SS	MEDS-26/A
Teacher in charge (if defined)	Luisa Barzon
Teaching Hours	10
Number of ECTS credits allocated	2
Course period	Second Semester
Course delivery method	<ul> <li>☑ In presence</li> <li>□ Remotely</li> <li>□ Blended</li> </ul>
Language of instruction	English
Mandatory attendance	<ul><li>☑ Yes (70% minimum of presence)</li><li>□ No</li></ul>
Course unit contents	In this course, the main characteristics and genetic and molecular bases of the pathogenesis of emerging and re-emerging viral infections will be explored in depth, with particular reference to those caused by pathogens identified as priorities by the WHO, such as Ebola virus, SARS-CoV-2, dengue virus, Zika virus, West Nile virus, and monkeypox virus. Additionally, the drivers determining the emergence of new pathogens (including climate change), epidemiological aspects, and methods of surveillance and early warning will be considered. The course will also cover prevention and control strategies, laboratory diagnosis, drugs, and vaccines in development. Case studies will be presented and discussed interactively.
Learning goals	English
Teaching methods	The aim of this course is to provide an in-depth understanding of the characteristics, genetic and molecular bases, epidemiology, and control strategies of emerging and re-emerging viral infections prioritized by the WHO, including interactive discussions of case studies.
Course on transversal, interdisciplinary, transdisciplinary skills	⊠ Yes □ No



Available for PhD students from other courses	⊠ Yes □ No
Prerequisites (not mandatory)	None
Examination methods (if applicable)	Not applicable
Suggested readings	Teaching materials provided by the instructor
Additional information	



Course unit English denomination	Big data analysis and bioinformatics	
SS	IBIO-01/A	
Teacher in charge (if defined)	Silvio Bicciato	
Teaching Hours	15	
Number of ECTS credits allocated	3	
Course period	03/2026	
Course delivery method	<ul> <li>☑ In presence</li> <li>□ Remotely</li> <li>□ Blended</li> </ul>	
Language of instruction	English	
Mandatory attendance	e ⊠ Yes 100% minimum of presence) □ No	
Course unit contents	<ol> <li>Introduction to the analysis of genomic data from high-throughput technologies high-throughput technologies for the analysis of genomic profiles         <ul> <li>online resources and databases for integrative genomics</li> <li>experimental design in the acquisition of genomic data with high-throughput technologies</li> <li>management of metadata and clinical information related to biological samples</li> </ul> </li> <li>Computational methods for the analysis of RNA-seq data         <ul> <li>quality control of raw data (raw reads), sequence alignment, and visualization of sequence alignments</li> <li>algorithms to quantify gene expression levels</li> <li>supervised analysis methods to identify phenotype markers of cells and tissues in different physiological states</li> <li>computational methods to infer cell fractions from bulk gene expression profiles (deconvolution methods</li> <li>computational strategies for the annotation and the functional interpretation of genomic data</li> </ul> </li> </ol>	



Learning goals	integrative analysis samples. Specific the analysis of ge and their integra evaluate the fur interaction networ	a to provide the knowledge and skills necessary for the sis of genomic data and phenotypic traits of biological c attention will be dedicated to computational methods for enomic profiles obtained with high-throughput technologies ation with the sample bio- molecular characteristics to nction of genes, regulatory sequences, and molecular ks. rning outcomes of the class are as follows:
	1. Knowle	dge and understanding
	a)	apply mathematical, statistical, and computer methods to the analysis of genomic data and phenotypic traits
	b)	use computational methods for genomic and post-genomic data analysis
	2. Ability t	o apply
	a)	logical-mathematical, statistical, and informatic tools
	b)	bioinformatics knowledge
	c)	bioinformatics techniques for the analysis, managemer and interpretation of genomic and bio-molecular data
	3. Autono	my of judgment
	a)	plan and interpret experimental data generated with high- throughput technologies
	b)	critically evaluate the computational and experimental methodologies for the integrated analysis of genomic data and molecular characteristics of biological samples
	C)	interpret the results of the analysis of genomic data in the light of the sample bio-molecular traits
Teaching methods	with the aid of IT the R computati expected to learn IT tools essentia bio-molecular data	elivered through face-to-face lectures which are carried out supports (Power Point presentations and data analysis in onal environment and in the Unix shell). Students are the theoretical and applicative aspects of algorithms and al for the integrative analysis of genomic profiles and a. The practical activities involve the application of methods analysis on case studies
Course on transversal, interdisciplinary, transdisciplinary skills	⊠ Yes □ No	
Available for PhD students from other courses	⊠ Yes □ No	



Prerequisites (not mandatory)	
Examination methods (if applicable)	
Suggested readings	<ul> <li>Lecture notes, bibliographic material, R scripts, and example genomic data</li> <li>B. Tijs, F. van der Kloet, M. Galland. RNA-seq lesson. Version 2020.04 (https://scienceparkstudygroup.github.io/rna-seq-lesson/index.html)</li> <li>A. Akalin. Computational genomics with R (https://compgenomr.github.io/book/)</li> <li>S. Holmes, W. Huber. Modern statistics for modern biology (https://web.stanford.edu/class/bios221/book/index.html)</li> </ul>
Additional information	





Course unit English	Functional interpretation of genomic data
denomination	
SS	IBIO-01/A
Teacher in charge (if defined)	Silvio Bicciato
Teaching Hours	15
Number of ECTS credits allocated	3
Course period	05/2026
Course delivery method	<ul> <li>☑ In presence</li> <li>□ Remotely</li> <li>□ Blended</li> </ul>
Language of instruction	English
Mandatory attendance	e ⊠ Yes (100% minimum of presence) □ No
Course unit contents	<ol> <li>Introduction to annotation and functional interpretation of Genomic profiling experiments from high-throughput technologies</li> <li>Gene sets for the functional annotation of genomic data         <ul> <li>construction of gene sets</li> <li>collections of molecular signatures and gene sets</li> </ul> </li> <li>Computational methods for annotation and functional interpretation of transcriptional and ChIP-seq profiles         <ul> <li>gene set enrichment methods: over-representation analysis</li> <li>gene set enrichment methods: functional class scoring</li> </ul> </li> </ol>
Learning goals	The course aims to provide the ability to use computational tools for the functional interpretation of genomic data and their analyses. The student will acquire fundamental concepts of computational methods for the functional annotation of results obtained from bioinformatic analyses of genomic data. Through its various components, the course seeks to develop the following skills:     1. Knowledge and understanding     a) in the application of mathematical, statistical, and computational methods for the integrated analysis of



				genomic data
			b)	in the use of computational methods for the functional interpretation of genomic data analysis
	2.	Ability	' to a	apply
			a)	logical-mathematical, statistical, and computational tools
			b)	bioinformatics knowledge
			c)	bioinformatics techniques for the functional interpretation of genomic data analysis
	3.	Indep	end	ent judgment
			b) me and c)	n evaluating and functionally interpreting genomic data in critically assessing computational and experimental thodologies that enable the integrated study of genomes d molecular characteristics of biological samples in interpreting the results of genomic data analysis in the ntext of the molecular mechanisms of cellular systems
Teaching methods	digital computa expecte computa The pra interpret	tools ational d to lea ational ctical a tation c	(Po env arn tool activ of ge	vered through in-person lectures, conducted with the aid of werPoint presentations and data analysis in the R vironment and with specific applications). Students are both the theoretical and practical aspects of algorithms and is essential for the integrated analysis of genomic profiles. vities involve the application of methods for the functional enomic data analysis results on case studies. A key aspect student's personal laptop during the lectures
Course on transversal interdisciplinary, transdisciplinary skills	'⊠ Yes □ No			
Available for PhD students from other courses	⊠ Yes □ No			
Prerequisites (not mandatory)	Big dat	a analy	/sis	and bioinformatics
Examination methods (if applicable)				
Suggested readings	Lecture genomic		bib	liographic material, R scripts, and example



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Additional information



Course unit English denomination	Gene editing: technologies and applications	
SS	BIO/17, BIO/11	
Teacher in charge (if defined)	Michelangelo Cordenonsi	
Teaching Hours	10	
Number of ECTS credits allocated	2	
Course period	May – June 2026	
Course delivery method	<ul> <li>☑ In presence</li> <li>□ Remotely</li> <li>□ Blended</li> </ul>	
Language of instruction	English	
Mandatory attendance	e ⊠ Yes 100% minimum of presence) □ No	
Course unit contents	Classic gene editing technologies: gene knockout and knockin in mouse embryonic stem cells. Conditional gene editing in mice: strategies and opportunities. Mouse breeding strategies to obtain complex genotypes. Gene editing technologies using CRISPR/Cas9: basic concepts. Application of CRISPR/Cas9 for gene knockout in human and murine cells: advantages and disadvantages. Targeted mutagenesis using CRISPR/Cas9. Gene knockin with CRISPR/Cas9 and its application to achieve conditional gene editing. Other conditional gene editing strategies with CRISPR/Cas9. Gene activation induced by defective-Cas9: opportunity for gain-of-function experiments. Screening using CRISPR/Cas9: loss-of-function and gain-of- function approaches.	
Learning goals	Learn the basics of CRIPR/Cas9 technologies and the opportunities they create for biomedical research. Learn to design targeting specific genes	
Teaching methods	The teaching is delivered through face-to-face lectures which are carried out with the aid of IT supports (Power Point presentations and online tools for planning the experiments). Students are expected to learn the theoretical and applicative aspects of gene editing via CRISPR/Cas9, and IT tools essential for planning the experiments. The practical activities involve the application of online tools for planning the experiments.	



Course on transversal interdisciplinary, transdisciplinary skills	' □ Yes ⊠ No
Available for PhD students from other courses	⊠ Yes □ No
Prerequisites (not mandatory)	Basic knowledge of molecular biology methods
Examination methods (if applicable)	Not applicable
Suggested readings	
Additional information	





Course unit English denomination	Flow cytometry: from basic principles to research applications		
SS	BIOS-12/A HUMAN ANATOMY		
Teacher in charge (if defined)	Di Liddo Rossa		
Teaching Hours	10		
Number of ECTS credits allocated	2		
Course period	February-June 2026		
Course delivery method	<ul> <li>☑ In presence</li> <li>□ Remotely</li> <li>□ Blended</li> </ul>		
Language of instruction	English/Italian		
Mandatory attendance	e ⊠ Yes (70% minimum of presence) □ No		
Course unit contents	The course aims to provide the knowledge and skills of basic flow cytometry topics and applications. In detail, the teaching course will have the following educational objectives: - understanding of the basic principles of fluorescence and light-particle interactions; - learning of the general concepts related to fluidics, optics, and electronic systems of flow cytometers; - knowledge of the different aspects of flow cytometry, including the quality control of both instrument and measurable parameters; - training in biological sample preparation and detection of cell targets (membrane, cytoplasmic, and nuclear), cell proliferation, cytotoxicity, apoptosis, and signaling pathways; - data analysis and graphical representation.		
Learning goals	<ul> <li>Learning of the basic concepts of flow cytometry;</li> <li>set up of flow cytometry analysis protocols;</li> <li>cell handling and staining;</li> <li>data acquisition and analysis</li> </ul>		



Teaching methods	The Program includes a part of frontal teaching followed by interactive exercises using different strategies for the presentation of flow cytometric data.
Course on transversal, interdisciplinary, transdisciplinary skills	r ⊠ Yes □ No
Available for PhD students from other courses	⊠ Yes □ No
Prerequisites (not mandatory)	
Examination methods (if applicable)	
Suggested readings	
Additional information	





Course unit English denomination	Computational analysis of single cell genomic data	
SS	BIOS-08/A	
Teacher in charge (if defined)	Mattia Forcato	
Teaching Hours	20	
Number of ECTS credits allocated	4	
Course period	April - may	
Course delivery method	<ul> <li>☑ In presence</li> <li>□ Remotely</li> <li>□ Blended</li> </ul>	
Language of instruction	English	
Mandatory attendance	e ⊠ Yes (100% minimum of presence) □ No	
Course unit contents	<ul> <li>Introduction to the analysis of single cell sequencing data: single cell technologies and biological sample preparation</li> </ul>	
	<ul> <li>experimental protocols for the acquisition of single cell genomic profiles: transcriptional (scRNA-seq), chromatin accessibility (scATAC-seq) and multimodal data</li> </ul>	
	<ul> <li>Analysis of single cell transcriptional (scRNA-seq) data: sequence alignment; quantification of the expression; identification, visualization and annotation of cell populations</li> </ul>	
	<ul> <li>Advanced methods for the analysis of single cell transcriptional profiles: integration of multiple datasets and multimodal data; study of cell-cell communications; analysis of trajectories</li> </ul>	
Learning goals	s The course aims to provide the knowledge and skills necessary for t integrative analysis of single cell genomic data. Specific attention will dedicated to computational methods for the analysis of genomic profil obtained with single cell high-throughput technologies and their application the reconstruction of cellular ecosystems in eukaryotic tissues.	
Teaching methods	The teaching is delivered through face-to-face lectures which are carried out with the aid of IT supports (Power Point presentations and data analysis in computational environments available via web browser). Students are expected to learn the theoretical and applicative aspects of algorithms and IT tools essential for the integrative analysis of single cell	





	genomic profiles. The practical activities involve the application of methods for single cell genomic data analysis on case studies
Course on transversal interdisciplinary, transdisciplinary skills	'⊠ Yes □ No
Available for PhD students from other courses	⊠ Yes □ No
Prerequisites (not mandatory)	
Examination methods (if applicable)	
Suggested readings	Amezquita et al. Orchestrating Single-Cell Analysis with Bioconductor (https://bioconductor.org/books/release/OSCA/)
Additional information	



Course unit English denomination	Applied Structural Bioinformatics
SS	BIOS-08/A
Teacher in charge (if defined)	Stefano Toppo
Teaching Hours	10
Number of ECTS credits allocated	2
Course period	October 2025
Course delivery method	<ul> <li>☑ In presence</li> <li>□ Remotely</li> <li>□ Blended</li> </ul>
Language of instruction	English
Mandatory attendance	<ul><li>☑ Yes (100% minimum of presence)</li><li>□ No</li></ul>
Course unit contents	Topics of structural bioinformatics will be addressed with a practical approach, starting from sequence data that will be analyzed in various ways. From multiple sequence alignment to molecular phylogenetics and the construction of a protein structural model where amino acid mutations will be mapped for comparison. The practical part will be based on sequences from SARS-CoV-2 haplotypes.
Learning goals	Students will learn to use some bioinformatics techniques for the analysis of structural variants on proteins.
Teaching methods	Practical activity at the computer lab after a theoretical introduction to the topics
Course on transversal interdisciplinary, transdisciplinary skills	r ⊠ Yes □ No
Available for PhD students from other courses	□ Yes ⊠ No



Prerequisites (not mandatory)	None
Examination methods (if applicable)	
Suggested readings	Materials provided during the course unit
Additional information	